SANGER SEQUENCING BEADS PURIFICATION

(Basically identical to Elkin *et al.*, Biotechniques 2002)

This assumes that you start with a 10 µl sequencing reaction in 0.2 ml PCR tubes.

- add 20 µl of BET solution, mix well by pipetting
- Incubate at room temperature for 10 minutes
- Place tubes a plate magnet for about 1 minute
- Remove the supernatant
- Add 30 µl of 70% ethanol
- Remove the ethanol (be careful to remove all of it)
- Air dry for 5-10 minutes
- Remove from the magnet and add 15 µl of milliQ water
- Incubate at room temperature for 5 minutes
- Place tubes on a plate magnet for 2 minutes
- Transfer 14 µl of water solution to new tubes

BET solution (2 ml, 100 cleanups)

- Ethanol 100%	1.6 ml
- milliQ water	190 µl
- Tetrat Ethylene Glycol	160 µl
- Sera-Mag SpeedBeads	50 µl

The original protocol does not say anything about washing the Sera-Mag beads before adding them to the BET solution. For good measure (to remove the sodium azide), I put the 50 µl on a magnet, remove the supernatant, was once with Tris 10 mM pH 8.0, and then re-suspend them in 50 µl of Tris 10 mM pH 8.0